## Photon Reception and Seeing Light in Squid Eye

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Synopsis: A review on retinal conformation change is given by the present paper. The retinal conformation change occurs by absorption of light. The wavelength of the absorbed light depends upon the structure of retinal. This problem is not so simple because the structures of various retinals are different to each other. Our data depends upon rhodopsin types, but the PDB data is limited. We do not yet have every type of rhodopsin in the PDB data for every condition. As the next step, we review previous literature on rhodopsin-based photoreceptor cells. There are many studies of the cells. Unfortunately the available papers are not in all the areas required. But we can nevertheless understand photreceptor cell function by the steps from absorbing the light to cell potential changes. Our ultimate goal is to find the reason why squids are easily caught only using a lamp. The image is simple: that is the squid can recognize the light at its eye. But we would like to understand the mechanism of light perception of squids. All facts regarding light perception of squids have not yet been resolved. There stands our contribution to understanding squid vision.

### 1. Introduction

There exist many studies about squid eyes and photon reception [1–35]. Photon reception has at its base the conformation change of retinal [1–35]. We have been studying the conformational change of retinal [61–69]. The molecule referred to as retinal exists in the rhodopsin protein found in the outer segment of rod cells of vertebrate photoreceptors and the rhabdomeres of most invertebrates including squids [35]. Many such cells are found in retinas of squids [35]. For convenience we will refer to all rhodopsin bearing photoreceptors as *rods*. A good review of the structure and function of squid eye can be seen the book "Squid as Experimental Animals". H. R. Saibil provided a chapter [19] for this book [35]. The cuttlefish is a class of squid [35–60]. We know that squid is the general name for a class of sea creatures [1–35]. The fishing of squid is usually at night using light provided by a lamp. Many squid are got in this manner. Interestingly the cuttlefish is caught by different manner.

Since almost all quid can detect light catching squid is the good fishing for boat crews. Squid can recognize the brightness of light, and lamp-light is better used for catching the squid at night. Then we can understand the development of Japanese squid fishing well. The mechanisms of light detection by squids have many studies [1–60]. The first step of light recognition is the conformation change of retinal. For this reason, we have been studying retinal, not only the conformation change of cis-trans structure, but also other things about retinal, using Protein Data Bank (PDB) data [61–68]. Retinal attaches to the amino acids of the rhodopsin protein [62–64]. The rhodopsin molecule is a transmembrane protein, and rhodopsin helices form a transmembrane ion channel [19]. The photo-activated rhodopsin binds and activates a G-protein. Thus we know that rhodopsin is the initiating protein of the photo-transduction cascade.

The retinal acts like a gate to conformation changes to rhodopsin. The molecular level func-

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tion of retinal and rhodopsin structure are well-understood via many studies of squid [1–35]. Our work should now step up to the much higher level of squid light recognition. For this purpose, a chapter given by Saibil [19] is better roundup to read. The retina of the squid eye contains many rod cells. The rod cells produce electrical potentials that ultimately governs squid brain function. The brain function decides the squid behavior as the response to light input.

We review photon absorption concentrating upon conformational changes of the retinal molecule. Section 2 summarizes this evidence and other related material. Section 3 summarizes light recognition by squids, with role of rod cells are summarized in Section 3. In Section 4, we summarize all the material and then we conclude that the squid behavior at the night to the lamplight of a boat is to the squid just like solar brightness.

# 2. Retinal conformation and Rhodopsin function

We have been studied the structure of squid retinal. The structure of retinal has the alternative form of trans-configuration and 11-cis configuration as shown by quantum orbital distributions [61]. Fine structural changes are caused by the structure of the cis-trans conformation of retinal driven by light absorption [61]. We also quantified that change in terms of the distance between nearest pairs of atoms of retinal calculated from PDB data [62]. We confirmed that the retinal ring part faces the amino acid with the ring part of Phenylalanine and tryptophan [63]. In the Mem KU paper [64], we explored vector analysis of carbons <sup>10</sup>C-<sup>12</sup>C of squid retinal. In a second round of vector analysis of squid retina, we presented each line between carbons of retinal as three dimension vectors and applied vector analysis [65]. The energy of wavelength for visible light was estimated as the squid absorbs in-coming lamp-light [66]. The wavelengths of Lamps have a continuous distribution across intensity. The exact distribution depends upon the lamp. We understand light in terms of energy [66]. The structure change of retinal by absorbing of the light was also considered [67]. It is difficult to estimate what intensity distribution should be taken for actual squid eyes so that we estimate the energy for visible light over the range from blue to red [67]. We further examined the PDB data using the RasMol software representation for viewing [68]. We published pictures of PDB data on RasMol views [68]. This concludes our studies on retinal structures.

Rhodopsin function is tightly dependent on the structure of the included retinal. Following Hargrave's lecture [69], the rhodopsin is the protein that interacts with other molecules—substrates, inhibitors, ions, nucleic acids, carbohydrates, lipids, and other proteins—to provide the ways in which they carry out their various functions. The rhodopsin molecules show many functions so we cannot summarize them briefly. The photon absorbing molecule is the retinal in the rhodopsin. The retinal cis-trans conformation change [61–68] effects the structure change of rhodopsin [69].

We show a schematic picture of rhodopsin in figure 1. There are many helices to form the rhodopsin protein. In figure 1, every helix is a coiled one. We denoted a line to connect the helix and that structure is determined by the amino acid residues. Rhodopsin is a transmembrane molecule containing n-pieces of rhodopsin in the outer segment of each rod cell. The rhodopsins exist on the disk membrane of outer segment of rod cell of vertebrates and in the tubular microvilli of invertebrate rhadomeres as in the squid. The structure of a vertebrate rod cell is schematically shown at

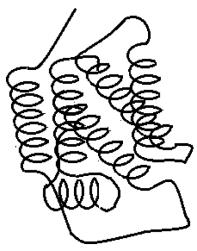


Fig. 1 schematic picture of rhodopsin

The coiled parts are helices with connected lines that consist of amino acids.

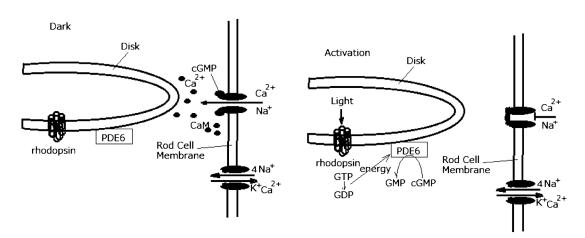


Fig. 2 Scheme for light activation of rhodopsin

#### figure 2.

The retinal is attached at the pore area of rhodopsin, but it is not shown in figure 1. Figure 1 is only a schematic illustration of a rhodopsin. The rhodopsin forms an ion channel path with gating proteins to realize the rhodopsin functions. Rhodopsin function is schematically understood by figure 2. Inward current of Ca<sup>2+</sup> and Na<sup>+</sup> is blocked by closed channel in brightness of light. Rhodopsin catalyzes GTP and releases energy by the chemical reaction (GTP->GDP+Pi+ energy). The energy comes from the third phosphate of GTP is catalyzed by rhodopsin protein. The energy transport thus occurs via absorption of light by the retinal molecule. The absorption drives the cis-trans conformation change of retinal. We know rhodopsin function from figure 2. We have no idea why rhodopsin exist on disks or microvilli. Invertebrate opsins are elongated and the microvilli thus constrain the rotation of the opsin causing the retinals to be somewhat aligned

making the microvilli dichroic. Orthogonal sets of microvilli allow squids and other animals to see linear polarization, which probably substitutes for color vision. Potential change is observed at rod cell as the voltage difference between inner and outer membrane of rod cell.

Rhodopsin is activated by light. Activated rhodopsin catalyze GTP to GDP so that this catalysis produce the decomposition energy from chemical reaction (GTP->GDP + Pi + energy). PDE 6 get this energy and opens on the cGMP to GMP. transition When rhodopsin is activated by light, inward current of Ca2+ and Na+ is stopped by the closed channel.

# 3. Rod cell and Light

Usually getting squid at night employs fish-boats and lights. Squid can recognize the incoming light by their eyes. The rod cells in the squid eye catches the light. Lights have an uneven distribution between wavelength and energy. The absorption of light is caused by the conformation change of retinal. The conformation change means structure change between trans-form and cis-form. Retinal binds to rhodopsin molecule at the inside of pore formed by rhodopsin on the disk mem-

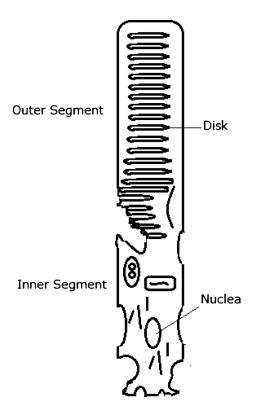
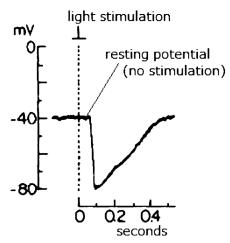


Fig. 3 Schematic Picture of Rod cell

The outer segment of a vertebrate rod cell, is occupied by many disks. Rhodopsin molecules exist in the membrane of disk of outer segment. In most invertebrate eyes the membranes are rolled into tubular microvilli. An exception are the day-time photoreceptors of scallops in which the photoreceptive membranes are rolled like paper towels where the axis about which the membranes are rolled pointing at the light. The scallop's night photoreceptors have microvilli like those of squid retinas.



**Fig. 4** Time course of potential change of rod cell after brief light stimulation

As seen the figure, a negative potential change appeared after light stimulation to the rod cell. from Dr. Janet Fizakerley homepage Copy right 2015 University of Minnesota Medical School Duluth, and we add a bit letters in the original figure.

brane. As seen from figure 3, rhodopsin exists on the outer segment of rod cell. The outer segment has a shell membrane and rhodopsin locate on the shell membrane from outside and inside of disk membrane with gating units. [35].

Following the homepage of Janet Fitazakerly at the University of Minnesota, the potential change occurs by light activation. Potential change against the time is illustrated by figure 4. If there is no light, the potential is a flat with small fluctuations (noise, mainly from failed intermittent stages in the catalytic cascade). The potential size depends somewhat upon the species, although are governed principally by the Nernst and Goldman equations for particular ion species. Figure 4 is seen when we chose one of retrieval results.

In the rod cell, conformation change of retinal by absorbing light is the first step of light activation. The retinal conformation change leads to structure change of rhodopsin. The retinal received the light energy and then that energy gates chemical energy stored in chemical bonds. This chemical energy induces the decomposition of GTP. The chemical reaction is written GTP->GDP + Pi + energy. The released energy makes the structure change: cGMP to GMP. This structure change is catalyzed by the enzyme phosphodiesterase (PDE6). This enzyme utilizes the released energy of GTP decomposition. The enzyme actively works in the brilliant conditions brought by lamp-light. The potential of rod cells is temporally changed to a more negative value as shown as figure 4. We only show potential change by light stimulation in the figure. Figure 4 says about a 40 mV change is seen against time development. This is the response of rod cell by light stimulation.

The rod cell function for absorbing the light has been summarized here. The details of the photochemical reaction were also reviewed in this section. Rod cells are found in the retina, and potential changes of the rod cells by light stimulation causes communication across retina. The potential change of the rod cells is transformed by other retinal cells and then reaches squid brain via action potentials, and the brain is controlled by those potential changes to produce a perception of the squid brain. The response of squid brain generates the behavior of the squid. The night behavior of

squid around the boats causes the squid fishing of the boats. The rod cell action by light is summarized in this section.

## 4. Summary and Discussion

The behavior of squid to see lights in the night is brought by a pathway, namely, retinal conformation change->rhodopsin function change->rod cell function->impulse movement of nerve cell->light recognition in the squid brain->response action of squid. This pathway of the squid runs within a very short time when the squid sees the light on the boat. Fishers know the squid behavior for the light and they get many squids. The behavior of squid is induced by rod cells in retina when the squid is seeing the light. The squid brain perceives brilliance or darkness, and squids come to brilliant light. Thus fisher gets the many squids using lamp-light. The behavior is different for squid or cuttlefish. The squid swims like a rocket with high speed, whereas the movement of cuttlefish is not high, namely it is not a rocket. Squids are fished at night using lamps and boats. Cuttlefishes are an alternative fishing target. In Japan, squids are used as the dried shredded ones. These dried squids are edible as foods. We know that Japanese people like the dried squid, so that owners of fish boats fishes the squids in Hokkaido prefecture because of easier fishing of squids compare than other areas.

One of the authors (Masashi Kito) has an interest squids, therefore he suggested that we write papers about squids. As stated above, we studied the retinal molecule in detail. But our overall goal is to understand what a squid is. The squid can perceive light, namely it understand dark and bright. Hence we should know the mechanism of understanding light in a squid. The pathway for understanding light is to know the root from light absorption through to the response behavior of squid. In the present paper, we consider rod cell functions. Since rod cell has the function of absorbing light and to mediate chemical reactions. As mentioned this paper, the required chemical energy is obtained by the decomposition of GTP (GuanineTriPhosphate). The required energy arises from the chemical bonding of the third phosphate. This may be written down as the chemical reaction GTP->GDP+Pi (phosphate)+energy. The chemical reaction needs water (chemical symbol is H<sub>2</sub>O). When we write the present paper, we know that a group of guaninephospate (GTP, GDP, GMP, cGMP) plays an important role to realize photon absorption to the potential change of rod cells. The function of rod cell is much the same for any species in as much as eyes have rod cells in retina. The rod cell potential change occurs by closing the channel to stop the inward current of Na<sup>+</sup> and Ca<sup>2+</sup>.

We should consider the further steps of the neurons connected with the rod cell, and ultimately the function of brain. The behavior of the squid to recognize the light appears to occur automatically. Fishers in boats can get many squids at the same time only using lamps and a fishing net. Extensive works about rod cells have been carried out so it is difficult to find new evidence. The PDB data of rhodopsin exist, and each PDB data contains retinal molecule data. The crystallized condition can be controlled so we know what configurations retinal molecules may undergo. We have therefore inferred information about retinal from the PDB data. We reviewed rod cell function and what things happen there. We may consider further visual processing steps work in the future.

#### References

- [1] J. E. Brown, D. C. C. C. Watkins, and C. C. Malbon, "Light-induced changes in the content of inositol phosphates in squid (Loligo pealei) retina", *Biochem. J.* **247** (1987) pp. 293–297.
- [2] A. I. Cohen, "An ultrastructural analysis of the photoreceptors of the squid and their synaptic connections. I. Photoreceptive and non-synaptic regions of the retina", J. Comp. Neurol. 147 (1973) pp. 351–378.
- [3] A. I. Cohen, "An ultrastructural analysis of the photoreceptors of the squid and their synaptic connections. II. Intraretinal synapses and plexus", J. Comp. Neurol. 147 (1973) pp. 379–398.
- [4] A. I. Cohen, "An ultrastructural analysis of the photoreceptors of the squid and their synaptic connections. III. Photoreceptor terminations in the optic lobes", J. Comp. Neurol. 147 (1973) pp. 399–426.
- [5] N. W. Daw, "Squid visual system", pp. 66-68, in: A guide to laboratory use of the squid Loligo pealei, (J. M. Arnold, W. C. Summers, D. L. Gilbert, R. S. Manalis, N. W. Daw, and R. J. Lasek, eds.), Marine Biological Laboratory, Woods Hole, Mass (1974).
- [6] N. W. Daw and A. L. Pearlman, "Pigment migration and adaptation in the eye of the squid, Loligo pealei", J. Gen. Physiol. 63 (1974) pp. 22-36.
- [7] F. M. Hagins, "Purification and partial characterization of the protein component of squid rhodopsin", J. Biol. Chem. 248 (1973) pp. 3298-3304.
- [8] W. A. Hagins, H. V. Zonana, and R. G. Adams, "Local membrane current in the outer segments of squid photoreceptors", *Nature* 194 (1962) pp. 844-847.
- [9] T. Hara and R. Hara, "Distribution of rhodopsin and retinochrome in the squid retina", J. Gen. Physiol. 67 (1976) pp. 791–805.
- [10] P. H. Hartline and G. D. Lange, "Optic nerve responses to visual stimuli in squid", *J. Comp. Physiol.* **93** (1974) pp. 37–54.
- [11] R. Hubbard and R. C. C. St. George "The rhodopsin system of the squid", J. Gen. Physiol. 41 (1958) pp. 501-528.
- [12] Y. Kito, T. Seki, T. Suzuki and I. Uchiyama, "3-dehydroretinal in the eye of a bioluminescent squid, *Watasenia scintillans*", *Vision Res.* **26** (1968) pp. 275–279.
- [13] G. D. Lange and P. H. Hartline "Retinal responses in squid and octopus", J. Comp. Physiol. 93 (1974) pp. 19-36.
- [14] S. Matsui, M. Seidou, I. Uchiyama, N. Sekiya, K. Hiraki, K. Yoshihara, and Y. Kito, 1988a, 4-Hydroxyretinal, a new visual pigment chromophore found in the bioluminescent squid, *Watasenia scintillans*", *Biochim. Biophys. Acta* **966** (1988) pp. 370–374.
- [15] K. Ozaki, R. Hara, T. Hara, and T. Kakitani, "Squid retinochrome. Configurational changes of the retinal chromophore", *Biophys. J.* 44 (1983) pp. 127–137.
- [16] Ozaki, K., Terakita, A., Hara, R., and Hara, T., 1987, Isolation and characterization of a retinal-binding protein from the squid retina, *Vision Res.* 27: 1057–1069.
- [17] L. H. Pinto and J. E. Brown, "Intracellular recordings from photoreceptors of the squid (*Loligo pealii*)", *J. Comp. Physiol.* 12 (1977) pp. 241–250.
- [18] C. F. E. Roper and K. J. Boss "The giant squid", Sci. Am. 246 (4) (1982) pp. 82-89
- [19] Helen R. Saibil "Structure and function of the squid eye", pp. 371–397 from Book "Squid as experimental animals" edited by D. L. Gilbert, W. J. Adelman Jr, and J. M. Aenold, (1990).
- [20] H. R. Saibil "An ordered membrane-cytoskeleton network in squid photoreceptor microvilli", J. Mol. Biol. 158 (1982) pp. 435-456.
- [21] H. Saibil "A light-stimulated increase of cyclic GMP in squid photoreceptors", FEBS Letters 168 (1984) pp. 213-216.
- [22] H. Saibil and E. Hewat "Ordered transmembrane and extracellular structure in squid photoreceptor microvilli", *J. Cell Biol.* **105** (1987) pp. 19–28.
- [23] H. Saibil and M. Michel-Villaz "Cross-reactions between rhodopsin and GTP-binding protein in bovine and squid photoreceptors", *Proc. Natl. Acad. Sci. USA* 81 (1984) pp. 5111-5115.
- [24] W. M. Saidel, J. Y. Lettvin, and E. F. MacNichol "Processing of polarized light by squid photoreceptors", *Nature* **304** (1983) pp. 534–536.
- [25] T. Seki, R. Hara, and T. Hara "Reconstitution of squid and cattle rhodopsin by the use of metaretinochrome in their respective membranes", Exp. Eye Res. 34 (1982) 609-621.
- [26] L. Sperling and R. Hubbard "Squid retinochrome", J. Gen. Physiol. 65 (1975) pp. 235-251.
- [27] I. Tasaki and T. Nakaye "Rapid mechanical responses of the dark-adapted squid retina to light pulses", *Science* 223 (1984) pp. 411-413.
- [28] S. I. Tomarev and R. D. Zinovieva "Squid major lens polypeptides are homologous to glutathione S-transferase subunits", *Nature* 336 (1988) pp. 86–88.
- [29] S. Tsukita, S., Tsukita, and G. Matsumoto "Light-induced structural changes of cytoskeleton in squid photoreceptor microvilli detected by rapid-freeze method", J. Cell Biol. 106 (1988) pp. 1151-1160.

- [30] C. A. Vandenberg and M. Montal "Light-regulated biochemical events in invertebrate photoreceptors. 1. Light-activated guanosine triphosphatase, guanine nucleotide binding, and cholera toxin catalyzed labelling of squid photoreceptor membranes", *Biochem.* 23 (1984) pp. 2339–2347.
- [31] C. A. Vandenberg and M. Montai "Light-regulated biochemical events in invertebrate photoreceptors. 2. Light regulated phosphorylation of rhodopsin and phosphoinositides in squid photoreceptor membranes", *Biochem.* 23 (1984) pp. 2347–2352.
- [32] C. A. Vandenberg and M. Montai, "Purification of squid rhodopsin and reassembly into lipid bilayers", *Biochim. Biophys. Acta* 771 (1984) pp. 74–81.
- [33] C. R. Worthington, S. K. Wang, and C. M. Folzer "Low angle X-ray diffraction patterns of squid *retina*", *Nature* **262** (1976) pp. 626-628.
- [34] H. V. Zonana "Fine structure of the squid retina", Johns Hopkins Hosp. Bull. 109 (1961) pp. 185-205.
- [35] D. L. Gilbert, W. J. Adelman Jr, and J. M. Aenold (eds) the book named "SQUID AS EXPERIMENTAL ANIMALS" Springer Sci + Business Media New York (1990).
- [36] J. J. Allen, L. M. Mathger, A. Barbosa, and R. T. Hanlon "Cuttlefish use visual cues to control three-dimensional skin papillae for camouflage." *J. Com. Physiol.* A195 (2009) pp. 547-555.
- [37] A. Barbosa, L. M. Mathger, K. C. Buresch, J. Kelly, C. Chubb, C., Chiao, and R. T. Hanlon, (2008). "Cuttlefish camouflage: The effects of substrate contrast and size in evoking uniform, mottle or disruptive body patterns", *Vision Res.* **48** (2008) pp. 1242–1253.
- [38] J. Bellingham, A. G. Morris, and D. M.Hunt (1998). The rhodopsin gene of the cuttlefish Sepia officinalis: sequence and spectral tuning. *J. Exp. Biol.* **201** (1978) pp. 2299-2306.
- [39] J. G. Boal, N. Shashar, M. M. Grable, K. H. Vaughan, E. R. Loew, and R. T. Hanlon, "Behavioral evidence for intraspecific signaling with achromatic and polarized light by cuttlefish (Mollusca; Cephalopoda)", *Behav.* **141** (2004) pp. 837–861.
- [40] J. Boal, R. A. Hylton, S. A Gonzalez, and R. T. Hanlon. "Effects of Crowding on the Social Behavior of Cuttlefish (Sepia officials)", Amer. Assoc. Lab. Animal Sci, No. 38 (1999): pp. 49-55.
- [41] B. B. Boycott "The functional organization of the brain of the cuttlefish Sepia officinalis", *Proc. R. Soc.* **153** (1961) pp. 503–534.
- [42] P. K. Brown and P. S. Brown "Visual pigments of the octopus and cuttlefish", Nature 182 (1958) pp. 1288-1290.
- [43] K. C. Buresch, L. M. Mathgar, J. J. Allen, C. Bennice, N. Smith, J. Schram, C. Chiao, C. Chubb, and R. T. Hanlon "The use of background matching vs. masquerade for camouflage in cuttlefish Sepia officinalis", *Vision Res.* 51 (2011) pp. 2362-2368.
- [44] C. Chiao, K. M. Ulmer, L. A. Siemann, K. C. Buresch. C. Chubb, and R. T. Hanlon "How visual edge features influence cuttlefish camouflage patterns". *Vision Res.* 83 (2013) pp. 40-47.
- [45] J. Forsythe, P. Lee, L. Walsh, and T. Clark. "The effects of crowding on growth of the European cuttlefish, Sepia officinalis Linnaeus, 1758 reared at two temperatures." J. Exp. Marine Biol. Eco., No. 269 (2002) pp. 173–185.
- [46] R. Hanlon, M.J. Naud, J. W. Forsythe, K. Hall, A. C. Watson, and J. McKechnie "Adaptable Night Camouflage by Cuttlefish." Am. Naturalist, No. 169 (2007) pp. 543–551
- [47] R. T. Hanlon and J. B. Messenger "Adaptive coloration in young cuttlefish (Sepia officinalis L.): The morphology and development of body patterns and their relation to behavior", *Phil. Trans. R. Soc. Lond.* B320 (1988) pp. 437– 487.
- [48] K. V. Langridge "Cuttlefish use start displays, but not against large predators", *Animal Behav.* 77 (2009) pp. 847–856.
- [49] L. M. Mathger, A. Barbosa, S. Miner, and R. T. Hanlon "Color blindness and contrast perception in cuttlefish (Sepia officinalis) determined by a visual sensorimotor assay", *Vision Res.* **46** (11) (2006) pp. 1746–53.
- [50] L. M. Mathger, R. T. Hanlon, J. Hakansson, and D. Nilsson "The W-shaped pupil in cuttlefish (*Sepia officinalis*): Functions for improving horizontal vision", *Vision Res.* 83 (2013) 19–24.
- [51] J. A. Miyan, T. Plymouth, R. T. Hanlon, and J. B. Messenger (1986). Short Commu in Neural Corr. C. Chan. Cuttlefish 400 (1986) 395–400.
- [52] M. Naud, and N. Jonathan "Sperm motility and longevity in the giant cuttlefish, *Sepia apama* (Mollusca: Cephalopoda)", *Marine Biology*, No. **148** (2006) pp. 559–566.
- [53] M. Norman, J. Finn, and T. Tregenza. "Female impersonation as an alternative reproductive strategy in giant cuttlefish." *The Royal Society*, No. **266** (1999) pp. 1347–1349.
- [54] B. Onsoy and A. Salman "Reproductive biology of the common cuttlefish *Sepia officinalis* L. (Sepiida: Cephalopoda) in the Aegean Sea", *Turkish J. Vete. Animal Sci.* **29** (2003) pp. 613–619.
- [55] M. Perez-Losada, A. Guerra, and A. Sanjuan "Allozyme differentiation in the cuttlefish *Sepia officinalis* (Mollusca: Cephalopoda)", NE Atla. Mediter. Heredity **83** (1999) pp. 280–289.
- [56] A. J. Shohet, R. J. Baddeley, J. C. Anderson, E. J. Kelman, and D. Osorio "Cuttlefish responses to visual orientation of substrates, water flow and a model of motion camouflage", J. Exp. Biology 209 (23) (2006) pp. 4717-4723

- [57] A. Sykes, P. M. Domingues, M. Correia, and J. P. Andrade. "Cuttlefish Culture- State of the Art and Future Trends." *Life & Enviro.*, No. **56** (2006) pp. 129–137.
- [58] Wang, J., G. J. Pierce, P. R. Boyle, V. Denis, J. P. Robin and J. M. Bellido "Spatial and temporal patterns of cuttlefish (*Sepia officinalis*) abundance and environmental influences—a case study using trawl fishery data in French Atlantic coastal, English Channel, and adjacent waters", ICES J. Marine Sci. 60 (5) (2003) pp. 1149–1158.
- [59] J. B. Wood "Cuttlefish husbandry. —Part IV; How do cuttlefish reproduce?—", Freshwater and Marine Aquarium Magazine 20 (10) (1998).
- [60] S. Zylinski, M. J. How, D. Osorio, R. T. Hanlon, and N. J. Marshall. "To Be Seen or to Hide: Visual Characteristics of Body Patterns for Camouflage and Communication in the Australian Giant Cuttlefish Sepia apama." THE AMERICAN NATURALIST, No. 177 (2011) pp. 681–690.
- [61] Y. Nagai, M. Kito, H. Kagawa, and T. Maddess. 2011 N0.32, "A quantum chemistry study of the retinal of squid rhodopsin", Mem. K. U. No. 32 (2011) pp. 20–31
- [62] 2 Y. Nagai, M. Kito, and T. Maddess "Spacing between retinal and amino acid residues in squid rhodopsin", *Mem K. U.* No. 33 (2012) pp. 1–7
- [63] Y. Nagai, M. Kito, and T. Maddess "Atomic configuration around retinal in squid rhodopsin", *Mem. K. U.* No. **34** (2013) pp. 1–8
- [64] Y. Nagai, M. Kito, and T. Maddess "Retinal location and structure in squid rhodopsin", *Mem. K. U.* No. 35 (2014) pp. 19–23
- [65] Y. Nagai, M. Kito, and T. Maddess. "A vector analysis of squid retinal", Mem. K. U. No. 36 (2015) pp. 1-7
- [66] Y. Nagai, M. Kito, and T. Maddess. "Estimation for energy transformation from retinal to rhodopsin in squid", Mem. K. U. No. 37 (2016) pp. 1-6
- [67] Y. Nagai, M. Kito, and T. Maddess "Energy absorption by structure change of squid retinal", Mem. K. U. No. 38 (2017) pp. 19-22
- [68] Y. Nagai, M. Kito, H. Wako, and T. Maddess "Analysis for cross-section of squid rhodopsin", Mem. K. U. No. 39 (2018) pp. 1-10
- [69] P. A. Hargrave, "Rhodopsin Structure, Function, and Topology", IOVS Vol. 42 (2001) pp. 3-9